

Whole-Genome Sequence of *Enteractinococcus helveticum* sp. nov. Strain UASWS1574 Isolated from Industrial Used Waters

Julien Crovadore,^a Gautier Calmin,^b Romain Chablais,^a Bastien Cochard,^a François Lefort^a

Plants and Pathogens Group, Research Institute Land Nature and Environment, hepia, HES-SO University of Applied Sciences and Arts Western Switzerland, Jussy, Geneva, Switzerland^a; Faculty of Engineering and Architecture, HES-SO University of Applied Sciences and Arts Western Switzerland, Delémont, Switzerland^b

We report here the whole-genome shotgun sequences of the strain UASWS1574 of the undescribed *Enteractinococcus helveticum* sp. nov., isolated from used water. This is the first genome registered for the whole genus.

Received 6 June 2016 Accepted 10 June 2016 Published 28 July 2016

Citation Crovadore J, Calmin G, Chablais R, Cochard B, Lefort F. 2016. Whole-genome sequence of *Enteractinococcus helveticum* sp. nov. strain UASWS1574 isolated from industrial used waters. *Genome Announc* 4(4):e00756-16. doi:10.1128/genomeA.00756-16.

Copyright © 2016 Crovadore et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to François Lefort, francois.lefort@hesge.ch.

Enteractinococcus is a new Gram-positive bacterial genus of the *Micrococcaceae* family created in 2012 (1) that contains four described species, *E. coprophilus* Cao et al. 2012, *E. fodinae* Cao et al. 2012 (formerly *Yaniella fodinae* Dhanjal et al. 2011), *Enteractinococcus lamae* Chen et al. 2015 (strain YIM 101617), and *E. viverae* Chen et al. 2015 (strain YIM 101632) (2), as well as three yet-undescribed species, including this strain. Mostly isolated from animal feces and soil, these bacteria are aerobic and nonmotile, coccoid to oval (0.5 to 1.5 μm diameter), and occur singly or in clusters. Growth was observed at 25 to 40°C (optimum 28°C) and at pH 7.0 to 11.0 (optimum pH 8.0); these species have a GC content in the range of 55.9 to 61.6% (1, 2).

The strain UASWS1574 was isolated from aerobic granules of industrial sewage sludge in an experiment of selection for highly ammonia-tolerant nitrifying bacteria. It was initially identified as belonging to the genus *Enteractinococcus* by 16S sequencing because it displayed 96 to 98% identity with the four known *Enteractinococcus* spp. Genomic DNA was extracted from a pure axenic culture grown to stationary phase following an adapted protocol (3). Libraries were generated using the TruSeq Nano DNA LT library kit (Illumina, USA). Whole-genome shotgun sequencing was carried out within one Illumina MiSeq run with 2 \times 250-bp paired-end read lengths, using the MiSeq reagent kit version 2 (Illumina) and providing a 114 \times genome coverage. Trimming and quality-control of the reads were performed with FastQC (4). Genome assembly was computed with SPAdes Genome assembler version 3.7.1 (5). The resulting contigs were arranged with BioEdit (6) and analyzed with QUAST (7). The final assembly yielded 118 contigs (≥ 200 bp.) with a total genome length of 3,670,653 bp, a GC content of 56.29%, and an N_{50} value of 176,870 bp.

Automated gene annotation was carried out by the NCBI Prokaryotic Genome Automatic Annotation Pipeline (PGAAP) (8) and reviewed with RAST version 2.0 (9). PlasmidFinder (10) did not detect any plasmid, which was confirmed by the RAST and PGAAP analyses. This bacterium owned 3,448 protein-coding sequences (CDSs) distributed in 362 subsystems, in which PGAAP identified 3,427 genes for 3,364 CDSs and 3,201 coding genes, 163

pseudogenes, and 63 RNA genes (5S, 16S, 23S, tRNAs, and ncRNAs). No complete transposon or phages were found integrated. The annotation confirmed the absence of toxins and superantigens, and virulence and disease genes were absent, therefore allowing this bacterium to be considered for industrial and environmental uses. The bacterium is equipped with resistance genes against metals such as arsenic, cadmium, chrome, cobalt, copper, mercury, and zinc and against a few antibiotics (penicillin, fluoroquinolones, and vancomycin). The bacterium is fully equipped for nitrate and nitrite ammonification, ammonia assimilation, and denitrification. With 60 genes involved in the metabolism of isoprenoids, this bacterium could be of interest for industry. One gene encodes a cyclohexene synthase, and two other enzymes are involved in cyclohexane and cyclohexanone degradation. Additionally, 68 genes are active in a wide variety of degradation pathways of aromatic compounds, thus offering a possible role in depollution of wastewater and contaminated soils.

Nucleotide sequence accession numbers. This whole-genome shotgun project was deposited at DDBJ/EMBL/GenBank under the accession number [LXEY00000000](https://www.ncbi.nlm.nih.gov/nuccore/LXEY00000000). The version described in this paper is the first version, LXEY00000000.1. The 118 contigs have been deposited under the accession numbers LXEY01000001 to LXEY01000118.

ACKNOWLEDGMENTS

This work was supported by the Swiss Federal Office for the Environment (FOEN) under grant number UTF 427.23.12, and by the Strategic Research Fund of the University of Applied Sciences and Arts Western Switzerland.

REFERENCES

1. Cao YR, Jiang Y, Jin RX, Han L, He WX, Li YL, Huang XS, Xue QH. 2012. *Enteractinococcus coprophilus* gen. nov., sp. nov., of the family *Micrococcaceae*, isolated from *Panthera tigris amoyensis* faeces, and transfer of *Yaniella fodinae* Dhanjal et al. 2011 to the genus *Enteractinococcus* as *Enteractinococcus fodinae* comb. nov. *Int J Syst Evol Microbiol* 62: 2710–2716. <http://dx.doi.org/10.1099/ijs.0.034249-0>.
2. Chen X, Li G-D, Li Q-Y, Hu C-J, Qiu S-M, Jiang Y, Jiang C-L, Han L,

- Huang X-S. 2015. *Enteractinococcus lamae* sp. nov. and *Enteractinococcus viverrae* sp. nov., isolated from animal faeces. *Antonie Van Leeuwenhoek* 108:1477–1483. <http://dx.doi.org/10.1007/s10482-015-0603-3>.
3. Lefort F, Douglas GC. 1999. An efficient micro-method of DNA isolation from mature leaves of four hardwood tree species *Acer*, *Fraxinus*, *Prunus* and *Quercus*. *Ann Sci* 56:259–263. <http://dx.doi.org/10.1051/forest:19990308>.
 4. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <http://www.bioinformatics.babraham.ac.uk/projects/fastqc>.
 5. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
 6. Hall TA. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucl Ac Symp Ser* 41:95–98.
 7. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUASt: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <http://dx.doi.org/10.1093/bioinformatics/btt086>.
 8. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Ciufu S, Li W. 2013. Prokaryotic genome annotation pipeline. In *The NCBI Handbook*, 2nd ed. NCBI, Bethesda, MD. <http://www.ncbi.nlm.nih.gov/books/NBK174280>.
 9. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75–90. <http://dx.doi.org/10.1186/1471-2164-9-75>.
 10. Carattoli A, Zankari E, García-Fernández A, Voldby Larsen M, Lund O, Villa L, Møller Aarestrup F, Hasman H. 2014. *In silico* detection and typing of plasmids using PlasmidFinder and plasmid multilocus sequence typing. *Antimicrob Agents Chemother* 58:3895–3903. <http://dx.doi.org/10.1128/AAC.02412-14>.