

AMERICAN SOCIETY FOR MICROBIOLOGY

Whole-Genome Sequences of Two *Arthrobacter* sp. Strains, 4041 and 4042, Potentially Usable in Agriculture and Environmental Depollution

Julien Crovadore,^a Damien Grizard,^b Romain Chablais,^a Bastien Cochard,^a Philippe Blanc,^c François Lefort^a

^aPlants and Pathogens Group, Research Institute, Land Nature and Environment, Hepia, HES-SO University of Applied Sciences and Arts Western Switzerland, Jussy, Geneva, Switzerland ^bProxis Développement, Levallois-Perret, France

^cBioprox, Noyant, France

ABSTRACT We report here the draft genome sequences of *Arthrobacter* sp. strains 4041 and 4042, both of which possibly belong to the diverse *Arthrobacter agilis* species and are potentially usable as plant biostimulants for agriculture and as depolluting bacteria for the environment.

A rethrobacter spp. (Actinobacteria) are Gram-positive soil bacteria, appearing in a rod or coccoid shape (1), which grow under aerobic and anaerobic conditions (1). They are present in Antarctic (2) and desert (3) soils and in alkaline and subglacial lakes (4, 5), and some species are known to promote plant growth (5–8), to inhibit plant-pathogenic bacteria and fungi or wood-decaying fungi (7, 9), and to degrade a wide range of organic and polyaromatic pollutants (4, 10, 11).

These two strains were isolated from soil samples in western France. DNA was extracted with a modified cetyltrimethylammonium bromide (CTAB) protocol (12) from a pure culture grown exponentially from a single colony in LB broth. The sequencing library was built with the TruSeg Nano DNA PCR-free library preparation kit (Illumina, USA). Whole-genome sequencing was carried out within one Illumina MiniSeq run at a 2 imes 151-bp paired-end read length using a MiniSeq high-output kit, with resulting genome coverages of $452 \times$ and $473 \times$ for strains 4041 and 4042, respectively. Overall quality metrics of the reads were assessed with FastQC version 0.11.5 (13). Genome assemblies were produced with SPAdes genome assembler version 3.10 (14), set in "paired-end assembly, careful mode," and yielded 31 and 34 contigs $(\geq 200 \text{ bp})$ for strains 4041 and 4042, respectively. They were finally ordered with BioEdit version 7.0.5 (15) and analyzed with QUAST version 4.6.3 (16) set as "QUAST: skip contigs shorter than 200 bp." The total genome length was 3,878,126 bp with a GC content of 67.66% and an N_{50} value of 466,984 bp for the strain Arthrobacter sp. 4041 and 3,235,327 bp with a GC content of 68.85% and an N_{50} value of 391,935 bp for the strain Arthrobacter sp. 4042. A BLAST search of the complete 16S rRNA gene of these 2 strains showed that these strains share about 99.7% identity with several Arthrobacter agilis strains in the GenBank nucleotide database (17). Automated gene annotation was carried out by the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) version 4.1 (18) and Rapid Annotations using Subsystems Technology (RAST) version 2.0 (19) using the ClassicRAST annotation scheme. PlasmidFinder version 1.3 (20) and plasmidSPAdes (21), both with default settings, did not detect any plasmids. PGAP identified 3,536 genes and 3,416 proteins in strain 4041 and 2,995 genes and 2,885 proteins in strain 4042. No known prophage was found. Based on the PGAP annotation, the NCBI genome neighbor report showed that strains 4041 and 4042 displayed 48.85% symmetric identity and

Received 28 July 2018 Accepted 20 August 2018 Published 13 September 2018

Citation Crovadore J, Grizard D, Chablais R, Cochard B, Blanc P, Lefort F. 2018. Wholegenome sequences of two *Arthrobacter* sp. strains, 4041 and 4042, potentially usable in agriculture and environmental depollution. Microbiol Resour Announc 7:e01054-18. https://doi.org/10.1128/MRA.01054-18.

Editor Julie C. Dunning Hotopp, University of Maryland School of Medicine

Copyright © 2018 Crovadore et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to François Lefort, francois.lefort@hesge.ch.

83.42% gapped identity with each other. Compared to the 2 other publicly available genomes, strain 4042 shared 99.85% symmetric identity and 99.99% gapped identity with *A. agilis* strain CGMCC 1.15723 from China, while 4041 displayed 56.82% symmetric identity and 86.29 gapped identity with strain UMCV2 from Mexico (8), confirming an observed high variability in the *Arthrobacter* genus (1). Their sequences also predicted resistance to antibiotics and toxic metal compounds. Strain 4041 has genes potentially involved in auxin synthesis and a nitrilase gene. Both strains are considered for agricultural and environmental uses.

Data availability. These whole-genome shotgun (WGS) projects were deposited at DDBJ/EMBL/GenBank under the accession numbers NFSC00000000 for *Arthrobacter* sp. strain 4041 and NFSD0000000 for *Arthrobacter* sp. strain 4042. The versions described in this paper are the first versions, NFSC01000000 and NFSD01000000. Concerning contigs, 31 and 34 contigs for *Arthrobacter* sp. strains 4041 and 4042, respectively, have been deposited at DDBJ/EMBL/GenBank under the accession numbers NFSC01000001 to NFSC01000031 and NFSD01000001 to NFSD01000034. Raw sequencing data sets have been registered in the NCBI Sequence Read Archive database (22) under the accession numbers SRR5513009 for strain 4041 and SRR5513012 for strain 2042.

ACKNOWLEDGMENTS

This work was supported by Cybèle Agrocare, Levallois-Perret, France, and the Strategic Research Fund of the University of Applied Sciences and Arts Western Switzerland.

REFERENCES

- Jones D, Keddie RM. 2006. The genus Arthrobacter. In Dworkin M, Falkow S, Rosenberg E, Schleifer KH, Stackebrandt E (ed), The prokaryotes. Springer, New York, NY. https://doi.org/10.1007/0-387-30743-5_36.
- Siebert J, Hirsch P. 1988. Characterization of 15 selected coccal bacteria isolated from Antarctic rock and soil samples from the McMurdo-Dry Valleys (South-Victoria Land). Polar Biol 9:37–44. https://doi.org/10 .1007/BF00441762.
- Eppard M, Krumbein WE, Koch C, Rhiel E, Staley JT, Stackebrandt E. 1996. Morphological, physiological, and molecular characterization of actinomycetes isolated from dry soil, rocks, and monument surfaces. Arch Microbiol 166:12–22. https://doi.org/10.1007/s002030050350.
- Kanekar PP, Sarnaik SS, Kelkar AS. 1998. Bioremediation of phenol by alkaliphilic bacteria isolated from alkaline lake of Lonar, India. J Appl Microbiol 85:1285–133S. https://doi.org/10.1111/j.1365-2672.1998 .tb05291.x.
- Singh RN, Gaba S, Yadav AN, Gaur P, Gulati S, Kaushik R, Saxena AK. 2016. First high quality draft genome sequence of a plant growth promoting and cold active enzyme producing psychrotrophic Arthrobacter agilis strain L77. Stand Genomic Sci 11:54. https://doi.org/10.1186/s40793-016 -0176-4.
- Velázquez-Becerra C, Macías-Rodríguez LI, López-Bucio J, Altamirano-Hernández J, Flores-Corte I, Valencia-Cantero E. 2011. A volatile organic compound analysis from *Arthrobacter agilis* identifies dimethylhexadecylamine, an amino-containing lipid modulating bacterial growth and *Medicago sativa* morphogenesis in vitro. Plant Soil 339:329–340. https:// doi.org/10.1007/s11104-010-0583-z.
- Velázquez-Becerra C, Macías-Rodríguez L, López-Bucio J, Flores-Cortez I, Santoyo G, Hernández-Soberano C, Valencia-Cantero E. 2013. The rhizobacterium Arthrobacter agilis produces dimethylhexadecylamine, a compound that inhibits growth of phytopathogenic fungi in vitro. Protoplasma 250:1251–1262. https://doi.org/10.1007/s00709 -013-0506-y.
- Valencia-Cantero E, Hernández-Calderón E, Velázquez-Becerra C, López-Meza LE, Alfaro-Cuevas R, López-Bucio J. 2007. Role of dissimilatory fermentative iron-reducing bacteria in Fe uptake by common bean (*Phaseolus vulgaris* L.) plants grown in alkaline soil. Plant Soil 291: 263–273. https://doi.org/10.1007/s11104-007-9191-y.
- Barrows-Broaddus J, Dwinell LD, Kerr TJ. 1985. Evaluation of Arthrobacter sp. for biological control of the pitch canker fungus (Fusarium monilisp. for biological control of the pitch canker fungus (Fusarium monilition)

forme var. subglutinans) on slash pines. Can J Microbiol 31:888-892. https://doi.org/10.1139/m85-166.

- Solyanikova IP, Emelyanova EV, Shumkova ES, Egorova DO, Korsakova ES, Plotnikova EG, Golovleva LA. 2015. Peculiarities of the degradation of benzoate and its chloro- and hydroxy-substituted analogs by actinobacteria. Int Biodeterior Biodegrad 100:155–164. https://doi.org/10.1016/j .ibiod.2015.02.028.
- Solyanikova IP, Suzina NE, Egozarian NS, Polivtseva VN, Prisyazhnaya NV, El-Registan GI, Mulyukin AL, Golovleva LA. 2017. The response of soil *Arthrobacter agilis* lush13 to changing conditions: transition between vegetative and dormant state. Environ Sci Health B 52:745–751. https:// doi.org/10.1080/03601234.2017.1356665.
- Lefort F, Douglas GC. 1999. An efficient micro-method of DNA isolation from mature leaves of four hardwood tree species Acer, Fraxinus, Prunus and Quercus. Ann Sci 56:259–263.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. http://www.bioinformatics.babraham.ac.uk/projects/ fastqc.
- Bankevich A, Nurk S, Antipov D, Gurevich A, Dvorkin M, Kulikov A, Lesin V, Nikolenko S, Pham S, Prjibelski A, Pyshkin A, Sirotkin A, Vyahhi N, Tesler G, Alekseyev M, Pevzner P. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Hall TA. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symp Ser 41:95–98.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https:// doi.org/10.1093/bioinformatics/btt086.
- Benson DA, Cavanaugh M, Clark K, Karsch-Mizrachi I, Lipman DJ, Ostell J, Sayers EW. 2013. GenBank. Nucleic Acids Res 41:36–42. https://doi .org/10.1093/nar/gks1195.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44: 6614–6624. https://doi.org/10.1093/nar/gkw569.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A,

A Microbiolog

Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75–90. https://doi.org/10.1186/1471-2164-9-75.

- Carattoli A, Zankari E, Garcia-Fernandez A, Voldby Larsen M, Lund O, Villa L, Aarestrup FM, Hasman H. 2014. PlasmidFinder and pMLST: in silico detection and typing of plasmids. Antimicrob Agents Chemother 58: 3895–3903. https://doi.org/10.1128/AAC.02412-14.
- Antipov D, Hartwick N, Shen M, Raiko M, Lapidus A, Pevzner P. 2016. plasmidSPAdes: assembling plasmids from whole genome sequencing data. Bioinformatics 32:3380–3387. https://doi.org/10.1093/bioinformatics/ btw493.
- 22. Leinonen R, Sugawara H, Shumway M. 2011. The Sequence Read Archive. Nucleic Acids Res 39:D19–D21. https://doi.org/10.1093/nar/gkq1019.