

Article

Implementation and Cross-Validation of a Pharmacokinetic Model for Precision Dosing of Busulfan in Hematopoietic Stem Cell Transplanted Children

Sylvain Goutelle ^{1,2,3} , Yann Thoma ⁴ , Roxane Buffet ^{5,6}, Michael Philippe ⁷, Thierry Buclin ⁸ ,
Monia Guidi ^{6,8,9}  and Chantal Csajka ^{4,5,9,*} 

- ¹ Hospices Civils de Lyon, Groupement Hospitalier Nord, Service de Pharmacie, 69005 Lyon, France
- ² Univ Lyon, Université Lyon 1, UMR CNRS 5558, Laboratoire de Biométrie et Biologie Evolutive, 69622 Villeurbanne, France
- ³ Univ Lyon, Université Lyon 1, ISPB, Faculté de Pharmacie de Lyon, 69008 Lyon, France
- ⁴ School of Management and Engineering Vaud (HEIG-VD), University of Applied Sciences and Arts Western Switzerland (HES-SO), 1401 Yverdon-les-Bains, Switzerland
- ⁵ School of Pharmaceutical Sciences, University of Geneva, 1205 Geneva, Switzerland
- ⁶ Institute of Pharmaceutical Sciences of Western Switzerland, University of Geneva, 1205 Geneva, Switzerland
- ⁷ Institut d'Hématologie et d'Oncologie Pédiatrique & Centre Léon Bérard, Département de Pharmacie, 69008 Lyon, France
- ⁸ Service and Laboratory of Clinical Pharmacology, Department of Laboratory Medicine and Pathology, Lausanne University Hospital and University of Lausanne, 1011 Lausanne, Switzerland
- ⁹ Center for Research and Innovation in Clinical Pharmaceutical Sciences, Lausanne University Hospital and University of Lausanne, 1011 Lausanne, Switzerland
- * Correspondence: chantal.csajka@chuv.ch; Tel.: +41-21-314-42-63



Citation: Goutelle, S.; Thoma, Y.; Buffet, R.; Philippe, M.; Buclin, T.; Guidi, M.; Csajka, C. Implementation and Cross-Validation of a Pharmacokinetic Model for Precision Dosing of Busulfan in Hematopoietic Stem Cell Transplanted Children. *Pharmaceutics* **2022**, *14*, 2107. <https://doi.org/10.3390/pharmaceutics14102107>

Academic Editor: Neal M. Davies

Received: 11 August 2022

Accepted: 22 September 2022

Published: 1 October 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Busulfan, a drug used in conditioning prior to hematopoietic stem cell transplantation (HSCT) in children, has a narrow therapeutic margin. The model-informed precision dosing (MIPD) of busulfan is desirable, but there is a lack of validated tools. The objective of this study was to implement and cross-validate a population pharmacokinetic (PK) model in the Tucuxi software for busulfan MIPD in HSCT children. A search of the literature was performed to identify candidate population PK models. The goodness of fit of three selected models was assessed in a dataset of 178 children by computing the mean error (ME) and root-mean-squared error of prediction (RMSE). The best model was implemented in Tucuxi. The individual predicted concentrations, the area under the concentration-time curve (AUC), and dosage requirements were compared between the Tucuxi model and a reference model available in the BestDose software in a subset of 61 children. The model from Paci et al. best fitted the data in the full dataset. In a subset of 61 patients, the predictive performance of Tucuxi and BestDose models was comparable with ME values of 6.4% and −2.5% and RMSE values of 11.4% and 13.6%, respectively. The agreement between the estimated AUC and the predicted dose was good, with 6.6% and 4.9% of the values being out of the 95% limits of agreement, respectively. To conclude, a PK model for busulfan MIPD was cross-validated and is now available in the Tucuxi software.

Keywords: busulfan; pediatrics; hematopoietic stem cell transplantation; oncology; population pharmacokinetics; therapeutic drug monitoring; model-informed precision dosing

1. Introduction

Hematopoietic stem cell transplantation (HSCT) is a procedure used for treating various hematological malignancies (e.g., leukemia) as well as the severe forms of nonmalignant hematological diseases such as sickle-cell anemia, thalassemia, aplastic anemia, and severe combined immunodeficiencies [1]. Busulfan is an alkylating agent widely used in conditioning regimens prior to HSCT in both adults and children.

The PK of intravenous busulfan has been extensively reviewed elsewhere [2,3]. Briefly, busulfan binds at a low extent to both the plasma proteins and erythrocytes. Its average volume of distribution is large, comparable to that of the total body water (0.7 L/kg). The metabolism of busulfan is complex. It primarily involves conjugation to glutathione by the glutathione-S-transferase (GST) enzymes in body cells. Renal elimination is a minor elimination pathway, with 30% of a dose excreted into the urine, mostly as metabolites (about 1% is excreted as an unchanged drug). Its half-life is short, ranging from 1.5 to 3 h in most individuals.

Busulfan is usually administered over 4 days, as an intravenous infusion every 6, 12, or 24 h, depending on the HSCT center's practice [4]. Busulfan has a narrow therapeutic margin. Its underexposure has been associated with graft rejection, while its overexposure has been associated with increased transplantation-related mortality [3,5,6]. An example of a potentially life-threatening complication of busulfan therapy is hepatic veno-occlusive disease.

The Busulfan area under the concentration-time curve (AUC) is the pharmacokinetic (PK) metric that best correlates with both the efficacy and toxicity of busulfan [5]. Busulfan's AUC target is expressed as either the cumulative AUC (AUC_{cum}) or the mean AUC per dose. The therapeutic drug monitoring (TDM) of busulfan has been recommended to determine drug exposure and adjust the drug dosage to avoid both under- and overexposure and to improve the outcome, especially in children [7–9]. The traditional method for estimating the AUC is based on rich sampling and noncompartmental analysis, but this method is laborious and uncomfortable for patients. An accurate estimation of busulfan's AUC through sparse sampling is possible when combined with model-based analysis [10]. This is usually performed via dedicated PK software programs that perform Bayesian estimation of individual PK parameters. This Bayesian approach requires a population PK model that provides prior information for PK parameter estimation. A number of population PK models of busulfan in HSCT children have been published in the last two decades [3]. However, only a few models have been implemented in clinical PK programs and evaluated for drug dosing in an external manner. Neely et al. have developed and validated a nonparametric model for the Bayesian dose adjustment of busulfan in children, which is available in the BestDoseTM software [10]. An AUC estimation based on only two blood samples was shown to be accurate compared with the AUC estimated from rich PK profiles. In addition, the dosage adjustments based on this model in an external cohort of patients were consistent with the dosage adjustments calculated using noncompartmental analysis in a reference laboratory in the USA. This model can thus be considered the best standard for the model-guided TDM and dose adjustment of busulfan in children. BestDose is the latest version of a program formerly known as USC*Pack, MM-USC*Pack, and RightDose. It has been developed for more than four decades by the Laboratory of Applied Pharmacokinetics and Bioinformatics (LAPKB), at the University of Southern California, Los Angeles, USA. It is available from the laboratory website (<http://www.lapkb.org/>) (accessed on 26 September 2022). More details are also available in a book edited by LAPKB directors [11]. However, BestDose only runs nonparametric PK models, which are scarce in population PK, as most published population models are parametric. Tucuxi is a new standalone program developed for the model-guided TDM of drugs [12]. It offers a user-friendly interface for clinicians, helping them in the process of drug dosage adjustment, by proposing concentration predictions, percentile calculations, and dosage adjustments. It has been developed by HEIG-VD in close collaboration with CHUV and is available via <http://www.tucuxi.ch> (accessed on 26 September 2022). The development team paid particular attention to user-friendliness and performance, to offer the best user experience to non-specialists. Its core computing engine is written in C++14, one of the best programming languages for efficient computation. Genericity is offered through the possibility to add new population PK models written in specific files, without modifying the software itself. Tucuxi only runs parametric PK models, but no busulfan model was available in the program so far. The objective of the present study was to implement and

cross-validate a PK model for the AUC estimation and dosage adjustment of busulfan in HSCT children with the Tucuxi software, using BestDose as a reference.

2. Materials and Methods

2.1. Literature Search for Candidate Population PK Models of Busulfan

First, we searched the literature to identify candidate busulfan population PK models for implementation in Tucuxi. We conducted a PubMed search in July 2018, using the keywords “busulfan” and “population pharmacokinetics”. The search period was restricted to 10 years (2008–2018). We screened the references of retrieved papers as well. We examined the title and abstract of the identified articles and discarded some of them based on the following exclusion criteria: adult population only, nonparametric model, and model including gene polymorphism as covariates. The latter criterion was justified because the genetic data on glutathione-S-transferase, the main metabolic pathway of busulfan, were not available in our patients’ cohort. Indeed, this test is not performed in routine clinical practice in most centers.

As it was not feasible to implement and cross-validate all the available models, we performed a second round of selection based on the following exclusion criteria: part of the population with age >20 years, number of patients <20, model with the number of compartments ≥ 2 , model not including body weight as a covariate, and reported without a description of all the necessary PK equations and parameters (e.g., fixed and random effects, residual error) for implementation in Tucuxi. The criteria related to the number of compartments and body weight covariate were justified by comparison with our reference model in BestDose.

2.2. Patients’ Data

We used a previously described dataset [13] to assess the predictive performance of the selected models and for subsequent analyses. Those data were collected as part of routine patient care in a hematology and oncology pediatric center in Lyon (Hospices Civils de Lyon, France), from June 2006 to June 2015. Ethics approval was obtained from two committees, including one from Hospices Civils de Lyon, as described previously [13]. We obtained a waiver of informed consent, as we only used the existing clinical data that were de-identified for this project.

Briefly, busulfan was administered over 4 consecutive days as a 2 h infusion administered every 6 h for 4 days (total of 16 doses) except in five patients (four were administered busulfan once daily and one twice daily). Busulfan TDM and dose individualization was performed for all the patients to achieve the conventional AUC target of 3.7–6.1 mg·h/L (900–1500 $\mu\text{M}\cdot\text{min}$) per dose, corresponding to an AUC_{cum} of 60 to 100 mg·h/L. Two blood samples were obtained at 0.5 and 2 h after the end of the first busulfan infusion in each patient. For some patients, TDM was repeated with the same two-sample collection on the subsequent days of therapy on the physician’s demand. A validated high-performance liquid chromatography (HPLC) assay was used to measure the busulfan concentrations. More details about the drug assay are available elsewhere [10].

The data from 178 individuals (85 males and 93 females) were available, with 491 measured busulfan concentrations. The median (min, max) values of age, body weight, and height were as follows: 4.0 (0.13–21) years, 16.7 (3.2–90.9) kg, and 103 (46–190) cm, respectively.

2.3. Goodness of Fit of Selected Models

The three models selected after the literature search were imported into the NONMEM[®] software. We created two versions of each model, with and without inter-occasion variability (IOV). Then, the models were used to fit the entire busulfan dataset by using the MAXEVAL = 0 option of NONMEM, which practically computes the individual Bayesian posterior parameter values based on both the prior information and the patients’ observations. We assessed the predictive performance of each model by computing the bias and

imprecision of each individual posterior model's predicted concentrations. The bias was defined as the mean percent error (MPE) of prediction:

$$MPE = \frac{1}{N} \sum_{i=1}^N \frac{C_{pred_i} - C_{obs_i}}{C_{obs_i}} \quad (1)$$

where C_{pred_i} and C_{obs_i} are individual predicted concentrations and observed concentrations, respectively.

The imprecision was defined as the root-mean-squared percent error (RMSE):

$$RMSE = \sqrt{\frac{1}{N} \sum_{i=1}^N \left(\frac{C_{pred_i} - C_{obs_i}}{C_{obs_i}} \right)^2} \quad (2)$$

The model providing the lowest values of bias and imprecision was considered the best model for our data, with imprecision being the primary criterion. The MPE and RMSE are classical criteria in the external validation of population PK models [14,15]. Diagnostic goodness-of-fit plots such as the individual predicted vs. observed concentrations, the conditional weighted residuals (CWRES) vs. population concentration predictions, and the time after dose further helped in assessing the most suitable model for our purpose.

2.4. Model Implementation in Tucuxi and Reference Model in BestDose

The model from Paci et al. [16] best fit the data in the NONMEM run (see the Results Section 3) and was imported into Tucuxi. This is a one-compartment model including the allometric scaling of busulfan clearance (CL) and the volume of distribution (V) to body weight (BW) as follows:

$$CL(L \cdot h^{-1}) = 2.18 \times \left(\frac{BW}{9} \right)^{\theta_1 \text{ or } \theta_2} \quad (3)$$

$$V(L) = BW^{\theta_3} \quad (4)$$

where the θ parameters are the allometric power coefficients. For CL, the coefficient value depends on the patient's weight, being $\theta_1 = 1.25$ in patients with $BW < 9$ kg and $\theta_2 = 0.76$ in patients with $BW \geq 9$ kg; for V, it is $\theta_3 = 0.86$. The model without IOV was imported in Tucuxi because IOV is currently not handled by the program.

This model was fit to individual patients' data in Tucuxi, and the results were compared with those obtained with our reference model, the nonparametric model from Neely et al. implemented in BestDose [10]. This is also a one-compartment model parameterized with an elimination rate constant (Ke) and V. The two parameters are allometrically scaled to BW or ideal body weight (IBW, estimated by the Traub–Johnson equation [17]), the lowest value being retained in each individual. The equations describing the covariate–parameter relationships are as follows:

$$Ke(h^{-1}) = Ke_S \times (BW \text{ or } IBW)^{-0.25} \quad (5)$$

$$V(L) = V_S \times (BW \text{ or } IBW) \quad (6)$$

In Equations (5) and (6), Ke_S and V_S represent the “slope” parameters, which are random. Notably, the model published by Neely et al. also included age as a covariate on both Ke and V, with a complex polynomial relationship. This relationship could not be included in the desktop version of BestDose, so only BW/IBW was retained. It has been shown that this simpler model provided bias and imprecision as good as those of the full model [10].

Table 1 summarizes the parameter values of the models implemented in Tucuxi and BestDose. Notably, for the nonparametric model (BestDose), the mean and standard deviation do not fully represent the parameter distribution, and therefore, the median

and minimal/maximal values are displayed. The typical values are reported for the parametric model.

Table 1. Parameters of the busulfan PK models implemented in Tucuxi and BestDose.

Tucuxi Model [16]		BestDose Model [10]	
Parameter	Value	Parameter	Value
Typical CL (L/h)	2.18	Median K_{ES} ($h^{-1} \cdot kg^{-0.25}$) (min–max)	0.71 (0.42–0.98)
Typical V (L)	$BW^{0.86}$	Median V_S ($L \cdot kg^{-1}$) (min–max)	0.72 (0.53–1.40)
CL variability (CV)	23%	K_{ES} variability (CV)	18%
V variability (CV)	22%	V_S variability (CV)	21%
Residual error	11% (proportional) 57 ng/mL (additive SD)	Residual error ^a	SD (mg/L) = 0.02 + 0.1·Cobs

^a In the residual error model, *Cobs* is measured busulfan concentration. Abbreviations: *BW*, body weight; *CL*, busulfan total body clearance; *CV*, coefficient of variation, *SD*, standard deviation.

2.5. Cross-Validation of Busulfan Model in Tucuxi

A probe subset of 61 individuals ($n = 119$ measured concentrations) who received busulfan every 6 h were randomly selected from the full dataset described above and used to compare the exposure and dosages predicted by the two models.

For each individual patient, the measured busulfan concentrations after the first dose were fitted with both the Tucuxi and BestDose models. The goodness of fit was assessed by computing the *MPE* and *RMSE* characterizing the comparison of observed concentrations and the individual posterior model predictions, as described previously. The AUC after the first dose (i.e., the first 6 h of therapy), denoted AUC_{0-6} , was computed with both models, as well as the predicted future dose required to achieve a cumulative AUC set at 80 mg·h/L for the entire therapy. In this prediction, we assumed that no intra-individual (inter-occasion) variability in the PK parameters would occur, so those parameters estimated after the first dose would hold unchanged for the entire therapy of 16 doses over 4 days.

To assess the correlation and agreement between the concentrations, the AUC, and the predicted doses from the two models, we performed linear regression and Bland–Altman analyses, respectively.

3. Results

3.1. Literature Search and Analysis

Fifty-four reports were initially retrieved from PubMed. We discarded 38 studies based on the primary exclusion criteria. After this first round, 16 studies were kept, and 4 other studies were identified from citations in the literature. A total of 20 reports were examined in the second round, and 17 were excluded based on the secondary exclusion criteria. In the end, three models were retained for the goodness-of-fit analysis. Those were the models from Paci et al. [16], Booth et al. [18], and Trame et al. [19].

3.2. Goodness of Fit of the Three Candidate Models for the Entire Dataset

Table 2 summarizes the characteristics of the three models and their predictive performance in the analysis of the entire dataset after NONMEM runs, with the corresponding goodness-of-fit plots shown in Supplemental Materials (Figure S1). For all the models, the predictive performance was improved when IOV was incorporated into the model. The model from Trame et al. showed the largest values of bias and imprecision. The models from Paci et al. and Booth et al. performed rather similarly regarding bias and imprecision. The model from Paci et al. was finally selected based on having the lowest imprecision and better *CWRES* diagnostic graphs than that of Booth et al. and then was imported into Tucuxi.

Table 2. Main characteristics of the selected models and predictive performance in the entire dataset ($n = 178$ patients, 491 measured concentrations).

Model [Reference]	Population Characteristics	Model Characteristics	MPE (%) [95% Confidence Interval]		RMSE (%)	
			Model without IOV	Model with IOV	Model without IOV	Model with IOV
Paci et al. [16]	$n = 205$ Median age, 2.5 years (10 days–15 years) Median BW, 12 (3.5–62.5) kg	One-compartment CL~BW (allometric) V~BW (allometric) IOV on CL	2.7 [1.7–3.7]	1.6 [0.8–2.3]	12.2	8.7
Booth et al. [18]	$n = 24$ Mean age, 6.3 years (3 months–16.7 years) Mean BW, 23.8 (7.1–62.6) kg	One-compartment CL~BW (allometric) V~BW (allometric) IOV on CL and V	2.1 [1.0–3.3]	1.1 [0.3–1.9]	13.4	9.5
Trame et al. [19]	$n = 94$ Median age, 9.2 (0.4–18.8) years Median BW, 27.2 (4.2–80) kg	One-compartment CL~BW (allometric) V~BW (allometric) IOV on CL and V	7.1 [5.1–9.3]	5.0 [3.2–6.8]	26.2	22.2

Abbreviations: BW, body weight; CL, busulfan clearance; IOV, inter-occasion variability; MPE, mean prediction error; RMSE, root-mean-squared error; V, busulfan volume of distribution.

3.3. Comparative Analysis of Tucuxi and BestDose results

Table 3 shows the predictive performance of the models in Tucuxi and BestDose in the 61 probe patients ($n = 119$ measured concentrations). Compared with the reference model in BestDose, the test model in Tucuxi showed a small but significant bias (+6.4%), indicating a slight overprediction of concentrations, with somewhat lower imprecision. The BestDose model displayed indicated a smaller but still significant negative bias (−2.5%). The results from Tucuxi were highly correlated and concordant with the results from BestDose regarding the predicted concentrations ($R^2 = 0.93$), the estimated AUC_{0-6} ($R^2 = 0.91$), and the predicted doses to achieve the target AUC ($R^2 = 0.97$), as shown in Figure 1.

Table 3. Predictive performance of the test and reference model in the individual analysis of 61 patients (119 measured concentrations).

Model (Software)	MPE (%) [95% Confidence Interval]	RMSE (%)
Test model (Tucuxi) [16]	6.4 [4.7; 8.1]	11.4
Reference model (BestDose) [10]	−2.5 [−0.3; −4.7]	13.6

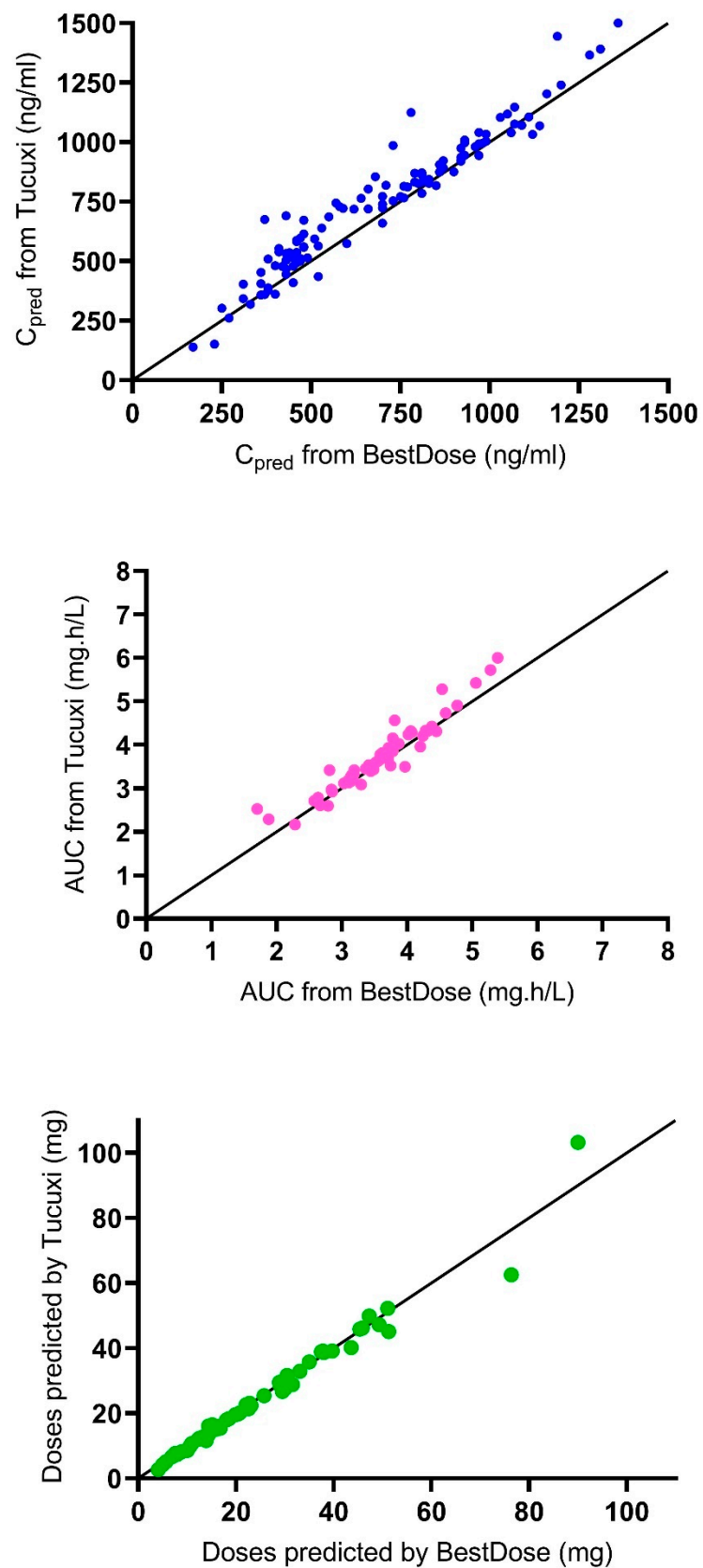


Figure 1. Correlation between results from Tucuxi and BestDose. Upper panel, predicted concentrations (C_{pred}); medium panel, estimated AUC_{0-6} ; bottom panel, predicted doses. The solid line is the line of identity ($y = x$).

Figure 2 represents the Bland–Altman analysis of the agreement between the results of the Tucuxi and BestDose models. For the predicted concentrations, the mean bias was $8.7 \pm 12.7\%$, and 6 out of 119 values (5%) were out of the 95% limits of agreement. For the estimated AUC_{0-6} , the mean bias was $3.7 \pm 7.4\%$, and 4 out of 61 values (6.6%) were out of the 95% limits of agreement. For the predicted dose required to achieve the target cumulative AUC of 80 mg·h/L, the mean bias was $-3.9 \pm 8.1\%$, and 3 out of 61 values (4.9%) were out of the 95% limits of agreement. Overall, this analysis indicated a fairly good agreement between the exposures and dosage requirements estimated by the two models.

Regarding the outlier results, one patient (male, 15 years old, 39 kg, 153 cm) exhibited low observed busulfan concentrations at 0.5 and 2.5 h post-dose (730 and 140 ng/mL, respectively), the latter being especially low. The predicted concentrations from Tucuxi and BestDose at 0.5/2.5 h post-dose were 691/151 ng/mL and 430/230 ng/mL, respectively. Therefore, in this patient, Tucuxi fitted the data quite well, while BestDose underestimated the first observations and overestimated the second one. This resulted in large percent differences on the Bland–Altman plot (+46.6% and -41.5%). As a result, the estimated AUC_{0-6} was low, and the estimates were not consistent between the two programs either, with a 38.8% difference on the Bland–Altman plot. Notably, this patient had TDM again on the second day of therapy with the observed concentrations of 750 and 540 ng/mL at 0.5 and 2.5 h post-dose, which suggests that an error might have occurred for the second measurement on the first day. Another large discrepancy in the predicted concentrations (difference of +58.4% on the Bland–Altman plot) was observed in a very young patient (female, 2 months old, 3.2 kg, 45.5 cm) who exhibited a high concentration 2.5 h post-dose (630 ng/mL), just slightly lower than the concentration 0.5 h post-dose (800 ng/mL), which suggests a very low clearance, probably due to immature metabolism. Again, this observation was better fitted by the Tucuxi software. In this patient, the dosage requirements predicted by the two programs were not consistent (-41.2% on the Bland–Altman plot).

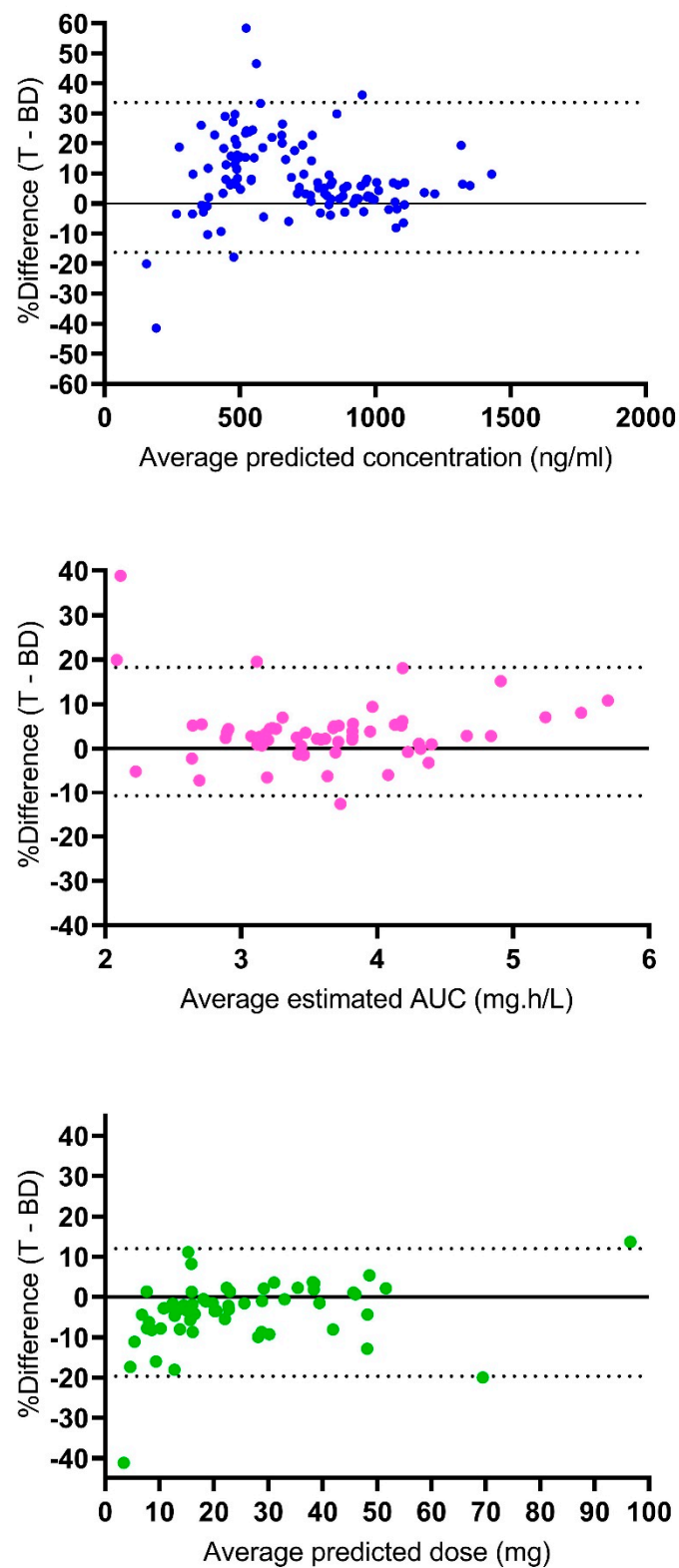


Figure 2. Bland–Altman plots of agreement between Tucuxi and BestDose results. Upper panel, agreement between predicted concentrations; medium panel, agreement between estimated AUC_{0–6}; lower panel, agreement between predicted doses. The dotted lines are the 95% limits of agreement. The solid horizontal line is $y = 0$.

4. Discussion

Busulfan population PK in children has been examined in numerous reports. In their recent review, Lawson et al. identified 21 population PK models of busulfan in children published between 2007 and 2019 [3]. Among those, a few models were externally validated in terms of the predicted concentrations. However, very few studies reported the implementation of such models in Bayesian PK programs and their validation for dose prediction, which are key for model-informed prediction dosing (MIPD). To our knowledge, the model from Neely et al. implemented in BestDose [10] is the only model that has been externally validated in terms of both drug exposure and dosage requirements.

We aimed at implementing a parametric population PK model in the Tucuxi software and cross-validate its results with those from the reference model of Neely et al.

Among the three models selected from the literature, the model from Paci et al. [16] best fit the entire dataset in the NONMEM run. This model was developed in a large number of patients, including a significant proportion of very young patients (33% of the patients with BW < 9 kg). The median age and weight were the closest to those of our cohort. The model from Booth et al. also performed well. The model from Trame et al. appeared less adequate. Compared with our dataset, this model was developed in older children, and part of them (57%) received oral busulfan. Those characteristics may explain its poorer performance when applied to our data.

The model from Paci et al. was then imported into the Tucuxi software and was individually fitted to the data from a random subset of 61 probe patients. The results were compared with those of the reference model from Neely et al. The predicted concentrations from Tucuxi showed a slight positive bias, while those from BestDose showed a very slight negative bias. As a result, the difference between Tucuxi and BestDose predictions was positive on average (see Figure 2). However, the agreement appeared well acceptable clinically. Importantly, both the estimated AUC_{0-6} and the predicted doses were highly correlated and concordant between the Tucuxi model and the reference model.

Both scatter plots and Bland–Altman plots showed that the differences between the two programs were larger for the predicted concentrations compared with the AUC or doses. There are several reasons explaining this. First, there were twice more data on concentrations, compared with the AUC and doses, yet more variability. Second, the prediction errors in drug concentrations may compensate, e.g., the concentration at 0.5 h may be underestimated, and that at 2.5 h may be overestimated. Finally, the error propagation from the concentrations to the AUC and the predicted dose is not linear and depends on the sampling time. The AUC depends mostly on the estimation of the CL parameter. The concentration obtained 2.5 h after the dose is more informative about CL than that obtained at 0.5 h post-dose (this one being more informative about busulfan V). Therefore, a given difference in the concentrations measured 0.5 h post-dose may result in a lower difference in the CL, AUC, and dose estimates between the programs.

Overall, our results confirm the adequacy of the model implemented in Tucuxi and validate its performance for busulfan dosing in children. Both models appear to be practically interchangeable for the AUC estimation and dosing decision.

The discrepancies described in two patients in the Results section illustrate underlying the differences in the model implemented in BestDose and Tucuxi. BestDose uses nonparametric Bayesian estimation, while Tucuxi uses a classical maximum a posteriori (MAP) Bayesian estimation. More details about the parametric and nonparametric approaches in population PK and TDM programs are available in the previous publications from our groups [20–22]. With the nonparametric Bayesian approach, probabilities are updated when fitting the data, but the parameter values from the discrete prior distribution do not change. This might result in poor fit in some outlier subjects because no set of parameter values among those available in the prior is adequate. By contrast, the MAP approach is more flexible, because the prior and posterior distributions of PK parameters are continuous. However, this flexibility may also be a weakness in case of a sampling or measurement error, because the estimation may converge, and the apparent good fit will

not warn users about the potential error. Strategies to cope with this, using, for example, the software-calculated percentiles to identify the out-of-scope measurements, are under investigation in Tucuxi. Notably, a hybrid fitting method not evaluated in the present study is available in BestDose and can better fit the outlier data [23].

Busulfan PK models are available in other dosing software tools, such as DoseMeRx, InsightRx, NextDose, and PrecisePK [24,25]. In a recent study, Lawson et al. compared the busulfan AUC predicted by models implemented in two different tools (NextDose and InsightRX) to the AUC calculated with the traditional trapezoidal rule, for various sampling strategies [26]. They showed that the bias and precision of the AUC estimation were not always consistent between software tools. However, they did not examine the agreement between the two PK programs. In addition, they only studied the AUC estimation, not the dosage requirements, which is arguably the most important criterion for MIPD. While the goodness of fit is a classic criterion in the validation of population PK models, we think that dose recommendations should be the primary diagnostic criterion when evaluating the dosing tools in a “fit-for-purpose” validation approach [27]. The previous studies evaluating dosing software considered this criterion as well [28,29].

This study has several limitations. The data were collected in routine patient care, and sampling for the drug’s concentration measurements was sparse. However, the reference model from Neely et al. was based on rich data and was externally validated based on part of this dataset [10]. The model from Paci et al. was also externally validated in the original study. Therefore, both models have been externally validated, and they are now cross-validated in the present study. We did not examine all the available models in our NONMEM analysis. Our primary goal was not to perform an external evaluation of all the published models but rather to identify a model to be cross-validated with our reference model implemented in BestDose. As our reference model was a one-compartment model, developed in children and adolescents only, including body weight and age as covariates, we preferred to select those models having similar characteristics to avoid confounders in the analysis of discrepancies. Model selection was based on both the goodness of fit and the characteristics facilitating the comparison with the reference model. Other models might be adequate as well and could be implemented in Tucuxi in the future. Indeed, it may be relevant to have several models available for a given drug in dosing software programs. This allows for a comparison of data fitting and dosage prediction for selecting the most adequate model for the population under consideration. It has been suggested that the best model should be chosen for each individual among a given set of competing models [28,30,31]. We did not evaluate the forecasting of concentration to be observed on subsequent TDM occasions, based on the first occasion data, because TDM was not repeated in all the patients. Although the models including IOV performed better in the NONMEM analysis, they could not be evaluated in the two dosing software programs, as neither currently handles IOV. By definition, IOV can be quantified a posteriori but is hardly predictable when designing a future dosage regimen. It seems that the best way to handle IOV in MIPD programs is to include IOV in the parameter estimation and data fitting but to ignore it in dose forecasting by considering IOV as an unexplained residual error [32,33]. Further research is necessary to confirm these findings.

To conclude, we implemented a PK model for busulfan dosing in children in the Tucuxi software. The model performance in terms of the estimated exposure and dosage prediction could be properly validated against a reference model available in BestDose. The results from both tools were consistent and indicated appropriate adequacy for the MIPD of busulfan in routine practice. Our approach may be proposed as a framework for the cross-validation of models used for MIPD in dosing software programs.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/pharmaceutics14102107/s1>, Figure S1: Goodness-of-fit plots of the candidate popPK models with (upper panel) and without (lower panel) interoccasion variability (IOV). Black continuous lines represent identity and ordinate value zero curves as appropriate, dashed lines the loess smooth curves. CWRES, conditional weighted residuals.

Author Contributions: Conceptualization, S.G. and C.C.; methodology, S.G., Y.T., R.B., M.G. and C.C.; software, S.G., M.P., Y.T., T.B. and C.C.; validation, S.G., Y.T., R.B. and C.C.; formal analysis, S.G., R.B. and C.C.; investigation, S.G. and M.P.; data curation, S.G. and M.P.; writing—original draft preparation, S.G. and R.B.; writing—review and editing, all authors.; supervision, S.G., Y.T. and C.C.; project administration, S.G. and C.C. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Ethics approval was obtained from two committees, including one from Hospices Civils de Lyon, as described previously [13]. The study was approved by the ethics committee of Hospices Civils de Lyon on 24 August 2017.

Informed Consent Statement: We obtained a waiver of informed consent, as we only used the existing clinical data that were de-identified for this project.

Data Availability Statement: Data are available upon request from the corresponding author.

Conflicts of Interest: The authors have no conflicts of interest that are relevant to the content of this study.

References

1. Hatzimichael, E.; Tuthill, M. Hematopoietic stem cell transplantation. *Stem Cells Cloning* **2010**, *3*, 105–117. [[CrossRef](#)] [[PubMed](#)]
2. Cremers, S.; Schoemaker, R.; Bredius, R.; Hartigh, J.D.; Ball, L.; Twiss, I.; Vermeij, P.; Vossen, J. Pharmacokinetics of intravenous busulfan in children prior to stem cell transplantation. *Br. J. Clin. Pharmacol.* **2002**, *53*, 386–389. [[CrossRef](#)] [[PubMed](#)]
3. Lawson, R.; Staatz, C.E.; Fraser, C.J.; Hennig, S. Review of the Pharmacokinetics and Pharmacodynamics of Intravenous Busulfan in Paediatric Patients. *Clin. Pharmacokinet.* **2021**, *60*, 17–51. [[CrossRef](#)] [[PubMed](#)]
4. Ruutu, T.; van der Werf, S.; van Biezen, A.; Backman, J.T.; Peczynski, C.; Kroger, N.; Mohty, M.; Nagler, A.; Montoto, S.; Langebrake, C.; et al. Use of busulfan in conditioning for allogeneic hematopoietic stem cell transplantation in adults: A survey by the Transplant Complications Working Party of the EBMT. *Bone Marrow Transplant.* **2019**, *54*, 2013–2019. [[CrossRef](#)] [[PubMed](#)]
5. Bartelink, I.H.; Lalmohamed, A.; van Reij, E.M.; Dvorak, C.C.; Savic, R.M.; Zwaveling, J.; Bredius, R.G.; Egberts, A.C.; Bierings, M.; Kletzel, M.; et al. Association of busulfan exposure with survival and toxicity after haemopoietic cell transplantation in children and young adults: A multicentre, retrospective cohort analysis. *Lancet Haematol.* **2016**, *3*, e526–e536. [[CrossRef](#)]
6. Feng, X.; Wu, Y.; Zhang, J.; Li, J.; Zhu, G.; Fan, D.; Yang, C.; Zhao, L. Busulfan systemic exposure and its relationship with efficacy and safety in hematopoietic stem cell transplantation in children: A meta-analysis. *BMC Pediatr.* **2020**, *20*, 176. [[CrossRef](#)]
7. Bleyzac, N.; Souillet, G.; Magron, P.; Janoly, A.; Martin, P.; Bertrand, Y.; Galambrun, C.; Dai, Q.; Maire, P.; Jelliffe, R.W.; et al. Improved clinical outcome of paediatric bone marrow recipients using a test dose and Bayesian pharmacokinetic individualization of busulfan dosage regimens. *Bone Marrow Transplant.* **2001**, *28*, 743–751. [[CrossRef](#)]
8. Philippe, M.; Goutelle, S.; Guitton, J.; Fonrose, X.; Bergeron, C.; Girard, P.; Bertrand, Y.; Bleyzac, N. Should busulfan therapeutic range be narrowed in pediatrics? Experience from a large cohort of hematopoietic stem cell transplant children. *Bone Marrow Transplant.* **2016**, *51*, 72–78. [[CrossRef](#)]
9. Palmer, J.; McCune, J.S.; Perales, M.A.; Marks, D.; Bubalo, J.; Mohty, M.; Wingard, J.R.; Paci, A.; Hassan, M.; Bredeson, C.; et al. Personalizing Busulfan-Based Conditioning: Considerations from the American Society for Blood and Marrow Transplantation Practice Guidelines Committee. *Biol. Blood Marrow Transplant. J. Am. Soc. Blood Marrow Transplant.* **2016**, *22*, 1915–1925. [[CrossRef](#)]
10. Neely, M.; Philippe, M.; Rushing, T.; Fu, X.; van Guilder, M.; Bayard, D.; Schumitzky, A.; Bleyzac, N.; Goutelle, S. Accurately Achieving Target Busulfan Exposure in Children and Adolescents With Very Limited Sampling and the BestDose Software. *Ther. Drug Monit.* **2016**, *38*, 332–342. [[CrossRef](#)]
11. Jelliffe, R.; Neely, M. *Individualized Drug Therapy for Patients. Basic Foundations, Relevant Software and Clinical Applications*; Elsevier: Amsterdam, The Netherlands, 2016; p. 434.
12. Dubovitskaya, A.; Buclin, T.; Schumacher, M.; Aberer, K.; Thoma, Y. TUCUXI: An Intelligent System for Personalized Medicine from Individualization of Treatments to Research Databases and Back. In Proceedings of the 8th ACM International Conference on Bioinformatics, Computational Biology and Health Informatics—ACM-BCB '17, Boston, MA, USA, 20–23 August 2017; pp. 223–232. [[CrossRef](#)]

13. Philippe, M.; Neely, M.; Rushing, T.; Bertrand, Y.; Bleyzac, N.; Goutelle, S. Maximal concentration of intravenous busulfan as a determinant of veno-occlusive disease: A pharmacokinetic-pharmacodynamic analysis in 293 hematopoietic stem cell transplanted children. *Bone Marrow Transplant.* **2019**, *54*, 448–457. [[CrossRef](#)] [[PubMed](#)]
14. Guo, T.; van Hest, R.M.; Roggeveen, L.F.; Fleuren, L.M.; Thorat, P.J.; Bosman, R.J.; van der Voort, P.H.J.; Girbes, A.R.J.; Mathot, R.A.A.; Elbers, P.W.G. External Evaluation of Population Pharmacokinetic Models of Vancomycin in Large Cohorts of Intensive Care Unit Patients. *Antimicrob. Agents Chemother.* **2019**, *63*, e02543-18. [[CrossRef](#)] [[PubMed](#)]
15. Zhao, C.Y.; Jiao, Z.; Mao, J.J.; Qiu, X.Y. External evaluation of published population pharmacokinetic models of tacrolimus in adult renal transplant recipients. *Br. J. Clin. Pharmacol.* **2016**, *81*, 891–907. [[CrossRef](#)] [[PubMed](#)]
16. Paci, A.; Vassal, G.; Moshous, D.; Dalle, J.H.; Bleyzac, N.; Neven, B.; Galambrun, C.; Kemmel, V.; Abdi, Z.D.; Broutin, S.; et al. Pharmacokinetic behavior and appraisal of intravenous busulfan dosing in infants and older children: The results of a population pharmacokinetic study from a large pediatric cohort undergoing hematopoietic stem-cell transplantation. *Ther. Drug Monit.* **2012**, *34*, 198–208. [[CrossRef](#)]
17. Traub, S.L.; Johnson, C.E. Comparison of methods of estimating creatinine clearance in children. *Am. J. Hosp. Pharm.* **1980**, *37*, 195–201. [[CrossRef](#)]
18. Booth, B.P.; Rahman, A.; Dagher, R.; Griebel, D.; Lennon, S.; Fuller, D.; Sahajwalla, C.; Mehta, M.; Gobburu, J.V. Population pharmacokinetic-based dosing of intravenous busulfan in pediatric patients. *J. Clin. Pharmacol.* **2007**, *47*, 101–111. [[CrossRef](#)]
19. Trame, M.N.; Bergstrand, M.; Karlsson, M.O.; Boos, J.; Hempel, G. Population pharmacokinetics of busulfan in children: Increased evidence for body surface area and allometric body weight dosing of busulfan in children. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **2011**, *17*, 6867–6877. [[CrossRef](#)]
20. Guidi, M.; Csajka, C.; Buclin, T. Parametric Approaches in Population Pharmacokinetics. *J. Clin. Pharmacol.* **2020**, *62*, 125–141. [[CrossRef](#)]
21. Goutelle, S.; Woillard, J.B.; Neely, M.; Yamada, W.; Bourguignon, L. Nonparametric Methods in Population Pharmacokinetics. *J. Clin. Pharmacol.* **2022**, *62*, 142–157. [[CrossRef](#)]
22. Goutelle, S.; Woillard, J.B.; Buclin, T.; Bourguignon, L.; Yamada, W.; Csajka, C.; Neely, M.; Guidi, M. Parametric and Nonparametric Methods in Population Pharmacokinetics: Experts' Discussion on Use, Strengths, and Limitations. *J. Clin. Pharmacol.* **2022**, *62*, 158–170. [[CrossRef](#)]
23. Goutelle, S.; Alloux, C.; Bourguignon, L.; Van Guilder, M.; Neely, M.; Maire, P. To Estimate or to Forecast? Lessons From a Comparative Analysis of Four Bayesian Fitting Methods Based on Nonparametric Models. *Ther. Drug Monit.* **2021**, *43*, 461–471. [[CrossRef](#)] [[PubMed](#)]
24. Kantasiripitak, W.; Van Daele, R.; Gijzen, M.; Ferrante, M.; Spriet, I.; Dreesen, E. Software Tools for Model-Informed Precision Dosing: How Well Do They Satisfy the Needs? *Front. Pharmacol.* **2020**, *11*, 620. [[CrossRef](#)] [[PubMed](#)]
25. Shukla, P.; Dvorak, C.C.; Long-Boyle, J.; Kharbanda, S. Lower Exposure to Busulfan Allows for Stable Engraftment of Donor Hematopoietic Stem Cells in Children with Mucopolysaccharidosis Type I: A Case Report of Four Patients. *Int. J. Mol. Sci.* **2020**, *21*, 5634. [[CrossRef](#)] [[PubMed](#)]
26. Lawson, R.; Paterson, L.; Fraser, C.J.; Hennig, S. Evaluation of two software using Bayesian methods for monitoring exposure and dosing once-daily intravenous busulfan in paediatric patients receiving haematopoietic stem cell transplantation. *Cancer Chemother. Pharmacol.* **2021**, *88*, 379–391. [[CrossRef](#)]
27. Keizer, R.J.; Ter Heine, R.; Frymoyer, A.; Lesko, L.J.; Mangat, R.; Goswami, S. Model-Informed Precision Dosing at the Bedside: Scientific Challenges and Opportunities. *CPT Pharmacomet. Syst. Pharmacol.* **2018**, *7*, 785–787. [[CrossRef](#)]
28. Burgard, M.; Sandaradura, I.; van Hal, S.J.; Stacey, S.; Hennig, S. Evaluation of Tobramycin Exposure Predictions in Three Bayesian Forecasting Programmes Compared with Current Clinical Practice in Children and Adults with Cystic Fibrosis. *Clin. Pharmacokinet.* **2018**, *57*, 1017–1027. [[CrossRef](#)]
29. Heitzmann, J.; Thoma, Y.; Bricca, R.; Gagnieu, M.C.; Leclerc, V.; Roux, S.; Conrad, A.; Ferry, T.; Goutelle, S. Implementation and Comparison of Two Pharmacometric Tools for Model-Based Therapeutic Drug Monitoring and Precision Dosing of Daptomycin. *Pharmaceutics* **2022**, *14*, 114. [[CrossRef](#)]
30. Goutelle, S.; Jay, L.; Boidin, C.; Cohen, S.; Bourguignon, L.; Bleyzac, N.; Wallet, F.; Vassal, O.; Friggeri, A. Pharmacokinetic/Pharmacodynamic Dosage Individualization of Cefepime in Critically Ill Patients: A Case Study. *Ther. Drug Monit.* **2021**, *43*, 451–454. [[CrossRef](#)]
31. Hughes, J.H.; Tong, D.M.H.; Lucas, S.S.; Faldasz, J.D.; Goswami, S.; Keizer, R.J. Continuous Learning in Model-Informed Precision Dosing: A Case Study in Pediatric Dosing of Vancomycin. *Clin. Pharmacol. Ther.* **2021**, *109*, 233–242. [[CrossRef](#)]
32. Abrantes, J.A.; Jonsson, S.; Karlsson, M.O.; Nielsen, E.I. Handling interoccasion variability in model-based dose individualization using therapeutic drug monitoring data. *Br. J. Clin. Pharmacol.* **2019**, *85*, 1326–1336. [[CrossRef](#)]
33. Keutzer, L.; Simonsson, U.S.H. Individualized Dosing With High Inter-Occasion Variability Is Correctly Handled With Model-Informed Precision Dosing—Using Rifampicin as an Example. *Front. Pharmacol.* **2020**, *11*, 794. [[CrossRef](#)] [[PubMed](#)]