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Artifacts in optical projection tomography due to refractive-index mismatch: model and correction

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Optical projection tomography (OPT) is a powerful tool for three-dimensional (3D) imaging of mesoscopic samples. While it is able to achieve resolution of a few tens of microns over a sample volume of several cubic centimeters, the reconstructed images often suffer from artifacts caused by inaccurate calibration. In this work, we focus on the refractive-index mismatch between the sample and the surrounding medium. We derive a 3D cone-beam forward model of OPT that approximates the effect of refractive-index mismatch. We then implement a fast and efficient reconstruction method to correct for the induced seagull-shaped artifacts on experimental images of fluorescent beads. © 2022 Optica Publishing Group

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Since its invention by Sharpe in 2002 [1,2], optical projection tomography (OPT) has become a powerful tool for obtaining three-dimensional (3D) images of biological tissues at the mesoscopic scale [3–5]. Due to its significant penetration depth [6], it can image whole animals at a resolution of a few tens of microns [4] with molecular specificity [3,4]. Often referred to as the optical analog of x ray computed tomography [7], OPT works with non-ionizing light, thereby minimizing radiation damage and facilitating studies of a broad range of diseases that include Alzheimer's [8] and gastrointestinal pathologies [4].

Regrettably, one frequently observes various artifacts in OPT images for a range of different reasons. For instance, mechanical errors in the imaging system may result in a misaligned rotation center that will generate double-edged or circular artifacts in the reconstruction [9–11]. Instabilities such as fluctuations of illumination and variations of the sensitivity of the detector will cause smear and ring artifacts [9]. Finally, deviations from the physical model may arise such as the finite depth-of-field of the imaging system [2] or a refraction that perturbs the propagation of light [6,12]. All these artifacts come from a mismatch between the OPT model and the actual physical realization. They strongly degrade the final resolution and quality of the reconstructed OPT images but can be computationally corrected once they have been properly identified [9,10].

During an OPT experiment, the sample is typically embedded in a cylindrical gel phantom that is immersed in an

index-matching liquid [2,13,14], in an attempt to limit the refraction of light at the interface of the agarose gel and the embedding liquid [13–16]. Despite their best effort, OPT practitioners still observe seagull-shaped artifacts suspected to be caused by a residual mismatch in the refractive indices (RI) between the sample and its embedding medium [4]. A previous study had already reported similar artifacts, mostly located around the edge of the sample, and proposed a computational method to compensate for the RI mismatch [6]. However, as this method requires volumetric alignment, it remains computationally expensive.

In this work, we propose instead a simple ray-optics model to remove artifacts induced by the RI mismatch between the cylindrical gel and the surrounding liquid. Our contributions are twofold: study of this type of RI mismatch and its interpretation as a virtual cylindrical lens; approximation of the virtual lensing effect with the help of a cone-beam model. We discuss the validity of our model and point out the importance of the displacement of the rotating cylinder. We then design a computationally efficient algorithm that leverages our model to correct the artifacts on experimental images of fluorescent beads. In an OPT experiment, the sample is rotated to a series of angular positions to produce a set of two-dimensional (2D) projections. Before the experiment, the rotation axis is carefully adjusted to be perpendicular to the optical axis, so that the projection data of one slice of the volume are recorded in a row of pixels on the detector [2]. When there is no refraction, diffraction, scattering, or defocusing, the emitted light propagates through the sample, the index-matching liquid, and the telecentric lens system in parallel straight lines (see Fig. 1). Accordingly, the standard forward model for OPT is the Radon transform [17,18], which in three-dimensions is typically inverted in a slice-by-slice fashion using the filtered backprojection (FBP) algorithm [2,17].

In reality, when the RI of the sample does not match the RI of the embedding liquid, the path along which the light propagates is altered so that the parallel-beam assumption no longer holds. Consider a 2D transverse slice of the sample in the liquid bath as depicted in Fig. 1. We denote the refractive index of the agarose gel and the liquid bath as n_1 and n_2 , respectively, and the radius of the cylinder as R . When $n_1 = n_2$, the light follows parallel straight lines as it travels through the sample, the liquid bath, and finally the flat wall of the cuvette (see the green lines in Fig. 1). When $n_1 < n_2$, the light will be refracted at the interface that separates

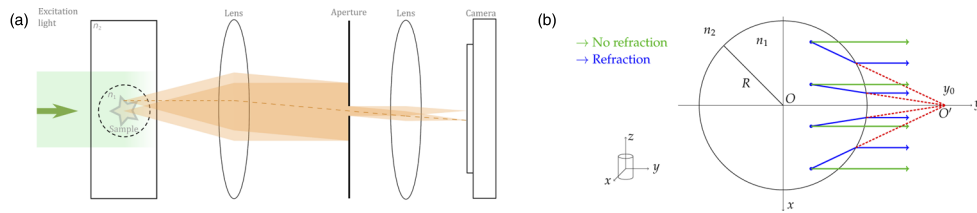


Fig. 1. OPT geometry in the ideal case and in the non-ideal case when the RIs do not match. (a) Emission OPT viewed from the top. The sample, fixed in agarose gel of RI n_1 , is placed inside a glass container filled with an embedding liquid of RI n_2 . The fluorophores inside the sample emit light that propagates through the object-space telecentric lens system and gets recorded by the camera. More details about the experimental system can be found in Ref. [4]. (a) Ideal case (green) and RI mismatch (blue). Only rays parallel to the optical axis are considered due to the low-numerical-aperture fluorescence collection in OPT. The RI n_1 of the agarose differs from the RI n_2 of the embedding medium. This creates a lensing effect, with a virtual focus at position O' on the optical axis.

the agarose cylinder from the liquid bath. The propagation of the refracted light is illustrated by the blue lines in Fig. 1. The extended incident rays (blue lines inside the cylinder in Fig. 1) intersect at a point O' on the optical axis (dotted lines in Fig. 1). Hence, the front wall (close to the detector side) of the cylinder can be thought of as a diverging lens whose virtual focus O' is on the optical axis. The rays inside the cylinder can then be approximated by a fan beam the origin of which coincides with the virtual focus.

Such a picture is based on two assumptions, common in ray optics, to keep our model computationally tractable. First, we make the paraxial assumption; namely, that the point emitters are close to the optical axis. Second, we consider the difference in RI to be small ($\Delta n = |n_2 - n_1| \ll n_1$), so that the resulting origin of the fan beam is far away from the cylinder. Under these assumptions, the location y_0 of the origin O' of the converging beam depends on the radius R of the cylinder and the RI (n_1, n_2) according to

$$y_0 = \frac{n_1}{n_2 - n_1} R. \quad (1)$$

A detailed derivation of Eq. (1) based on the ray geometry of Fig. 1 is provided in Supplement 1. Note that the dimensionless quantity y_0/R is not affected by the magnification of the imaging system.

Therefore, when $n_1 \neq n_2$, this physical model differs from the one typically used for reconstruction with a parallel-beam geometry. Yet, the proposed virtual-lens model is a better approximation of the actual physics. We are going to show that it is capable of reproducing and eventually correcting for some of the reconstruction artifacts. This 2D analysis corresponds to a fan-beam model; we can extend it to a cone-beam model when the height of the volume is sufficiently small. Cone-beam geometries are common in other tomographic applications, which allows us to leverage efficient implementations and reconstruction algorithms.

While Eq. (1) provides a good approximation of the beam origin in an idealized setting, two additional challenges arise in practice. First, the RI of the agarose gel evolves to match that of the surrounding liquid (e.g., BABB) due to molecular diffusion. This process may extend over the standard clearing protocol time, making it difficult to measure the value of n_1 and thus to calculate the origin using Eq. (1). Second, there is another physical effect to consider: the cylinder is displaced as it rotates because of an experimental offset between the axis of rotation and the axis of the cylinder. Despite their best effort, instrumentalists have not yet been able to calibrate their setups accurately enough to avoid this offset [18–20]. For example, the

seagull artifacts in Ref. [4] [cf. Fig. 2(f) therein] can be reduced by decreasing the offset between the axis of the cylinder and the axis of rotation. In Supplement 1, we show that the cone-beam geometry remains a good approximation under these conditions, but with an origin that is closer to the sample than Eq. (1).

To tackle both challenges at once, we propose a method to automatically determine the optimal origin of the cone beam by minimizing an heuristic loss metric. More details are provided in the section presenting experiments on real data. To validate our model of the RI mismatch, we simulate the setup of a real emission OPT for the imaging of fluorescent beads. We use the cone-beam model to generate projections with an origin position y_0 defined by $y_0/R = 5$. We then reconstruct with the FBP algorithm. Our phantom consists of 13 beads, each of unit intensity and of size (5×5) px (one pixel is 2.45 μm) embedded in a slice of a cylinder with radius $R = 1024$ px contained in a square cuvette of length 2048 px. Moreover, it is important to consider that the depth of field in OPT usually covers only half of the sample, in contrast with x ray tomography. To approximate the out-of-focus phenomenon of the other half, we implement rotation angles over both $[0, \pi)$ and $[0, 2\pi)$.

All simulations have been performed using the Tomosipo toolbox [21], which is a wrapper for the tomographic-reconstruction library ASTRA [22]. We have adapted a cone-beam geometry as this modality is common in tomography [23] and has already been implemented efficiently in Tomosipo. It enables us to accelerate the computations on central processing units (CPU) and graphics processing units (GPU) and to use the already implemented Feldkamp–Davis–Kress (FDK) algorithm for cone-beam reconstructions [24] in stacks of slices of size 2 to obtain the 3D volume. This is crucial in OPT because a time- and memory-efficient reconstruction algorithm is required to perform the high-resolution imaging of mesoscopic samples and to process huge datasets (1200 projections of (2048×2048) px each for the experiment presented below). Accurate models such as Ref. [6] hit computational bottlenecks and become infeasible for temporal and spatial imaging at high-resolution.

We show in Fig. 2(a) a vertical section of the reconstruction of a bead sitting at the exact center of the image. This ground-truth image is free of artifacts. As the distance of the bead to the center of the image increases, the seagull artifacts become more pronounced, which is reflected in the size of their “wings”. We give a close-up view of a non-centered bead with strong seagull artifacts in Figs. 2(b) and 2(c). More precisely, its reconstructions using the rotation angles in $[0, \pi)$ and $[0, 2\pi)$ are shown in Figs. 2(b) and 2(c), respectively. The magnified ground truth image is provided in Fig. 2(a) as a reference. In agreement

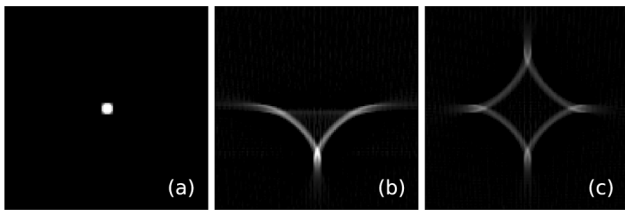


Fig. 2. “Seagull” artifacts due to mismatched RI with rotation angles $[0, \pi]$ (b) and $[0, 2\pi]$ (c), compared with the ground-truth fluorescent bead (a).

with our model, the FBP-based reconstruction of a mismatched experiment introduces similar artifacts to those observed in real data.

Next, we show how to mitigate the “seagull” artifacts on experimental data of fluorescent beads. Fluorescent FITC-labeled 1 m microspheres based on melamine resin (90305-5ML-F, Sigma) were used as specimen, in accordance with the preparation procedure described in Ref. [4]. Approximately 0.01 mL of particle solution was mixed to 10 mL of 1.5 % agarose before molding to achieve a nearly colloidal solution. The acquisition was performed shortly after preparation to avoid fluorescence quenching of the signal. The RI n_2 of BABB in our experiment is 1.56. The RI n_1 of the agarose changes over time, but the mismatch between the two materials is initially of the order of 0.1.

Before reconstruction, we first compensate for the misaligned rotation center. However, because the fluorescent beads are very sparse compared with an actual biological sample, they carry too little information for one to apply the standard method based on the maximum variance of a reconstructed slice [4,11,19]. Instead, we perform a grid search and determine the best estimate of the center of rotation when the “circle”-shaped artifacts caused by the wrong rotation center vanish.

As discussed before, another step is to determine the optimal origin of the cone beam. We do this automatically by minimizing the loss metric

$$E_{\text{loss}} = 1 - \frac{\sum_{k \in \Omega_{\text{in}}} I_k}{\sum_{k \in \Omega_{\text{total}}} I_k}, \quad (2)$$

where Ω_{total} is a square region that fully encloses the whole “seagull” artifact, Ω_{in} is a smaller square region enclosing only the central part of a bead [see Figs. 3(b), 3(d), and 3(f)], and I_k denotes the intensity at pixel k . The quantity E_{loss} hence indicates how concentrated the distribution of intensity is around the center of a bead. The smaller E_{loss} is, the less energy is spread outside the true location of the bead, and the better the reconstruction is. We provide in Fig. 3 the energy loss of five different beads at varied locations in the field of view. We observe that the energy loss behaves similarly for all five beads and achieves its minimum at approximately the same value of each curve. This means that the optimal origin guesses are consistent over several beads, which confirms the robustness of the proposed automatic calibration. We thus adopt the average value $\mu = y_0/R = 4.4$ as the optimal origin y_0 for all the beads in the reconstruction. Compared with μ , the theoretical value of $y_0/R = 15$ derived from Eq. (1) highlights the importance of performing this automatic calibration to account for the origin shift discussed in Supplement 1. The 3D reconstruction of a $2048 \times 2048 \times 1024$ volume and grid search to find the optimal origin over 200 points for one bead takes 95 seconds and 108 seconds on CPU and GPU, respectively.

In Fig. 3, we compare the reconstructions using the parallel-beam model or our approach. We can observe the clear radial dependence of the size of the seagull artifacts. Our proposed cone-beam model successfully mitigates the artifacts, especially for the beads identified by boxes. To quantify the improvement of our method, we provide the intensity profiles along the direction of maximal variance of the uncorrected and corrected

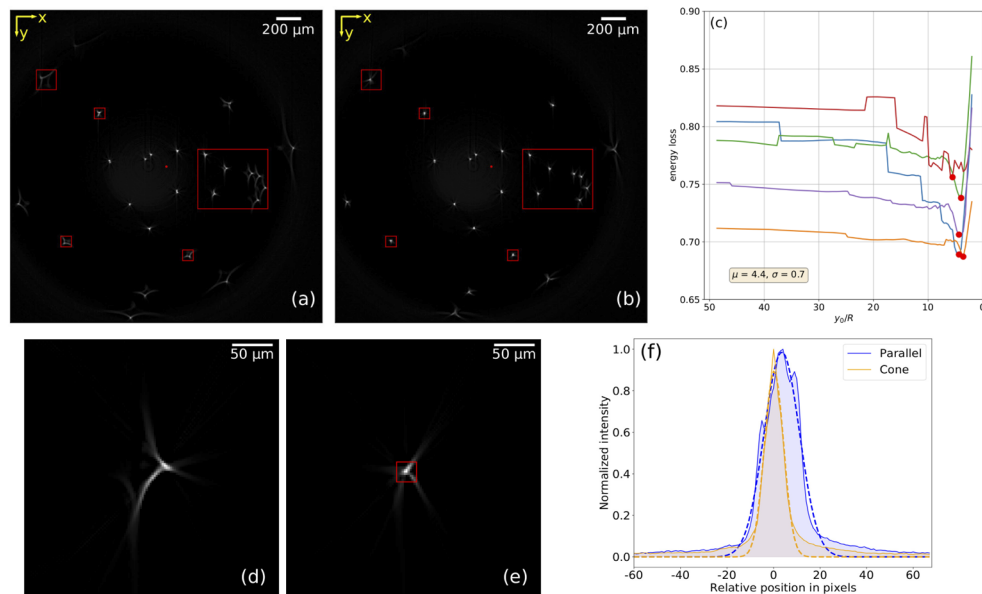


Fig. 3. Reconstructions of a 3D volume of fluorescent beads integrated along the z -axis using (a) a parallel-beam model and (b) a cone-beam model. (c) Energy loss of five beads over a range of origin guesses. The red point on each curve indicates the minimum. Same slice of the 3D reconstructions of a bead using (d) parallel-beam and (e) cone-beam models. The square in panel (e) is the region Ω_{in} used to compute the energy loss. (f) Intensity profiles along the direction of maximum variance of the bead in panels (d) and (e). The blue and orange lines represent the intensity profiles for the parallel-beam and cone-beam models, respectively. The dashed blue and orange lines represent the fitted Gaussian point-spread functions.

reconstructions in Fig. 3(f). The full-width half-maximum along the direction of maximum variance, calculated based on the Gaussian fit, is 47 μm and 22 μm for Figs. 3(d) and 3(e), respectively. This means that our corrections lead to a 52 % increase in resolution.

To conclude, this work models mismatching RI in emission OPT as a virtual lens. Using the corresponding cone-beam model, we can reproduce in simulations the “seagull” artifacts observed in real emission OPT experiments. To achieve best performance in practice, we propose a fast approach that automatically determines the optimal origin of the cone beam. We perform reconstructions based on the cone-beam model and validate our methods on experimental data of fluorescent beads. Our results show that the cone-beam model, with an optimized origin, successfully mitigates the seagull-shaped artifacts and increases the resolution of the reconstruction in the directions perpendicular to the optical axis. Future research directions include the design of a better model of the finite depth-of-field and of the rotating cylinder. This work could be extended to transmission OPT by taking into account the RI mismatch on both sides of the sample. Finally, cone-beam reconstructions could be applied to non-pointwise samples.

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Disclosures. The authors declare no conflicts of interest.

Data availability. Data underlying the results presented in this paper are available in [25].

Supplemental document. See Supplement 1 for supporting content.

REFERENCES

1. J. Sharpe, U. Ahlgren, P. Perry, B. Hill, A. Ross, J. Hecksher-Sørensen, R. Baldock, and D. Davidson, *Science* **296**, 541 (2002).
2. J. Sharpe, *Annu. Rev. Biomed. Eng.* **6**, 209 (2004).
3. A. Darrell, H. Meyer, U. Birk, K. Marias, M. Brady, and J. Ripoll, *2008 8th IEEE International Conference on Bioinformatics and BioEngineering*, (2008), pp. 1–6.
4. C. Schmidt, A. L. Planchette, D. Nguyen, G. Giardina, Y. Neuen-schwander, M. D. Franco, A. Mylonas, A. C. Descloux, E. Pomarico, A. Radenovic, and J. Extermann, *Biomed. Opt. Express* **12**, 3619 (2021).
5. D. Nguyen, P. J. Marchand, A. L. Planchette, J. Nilsson, M. Sison, J. Extermann, A. Lopez, M. Sylwestrzak, J. Sordet-Dessimoz, A. Schmidt-Christensen, D. Holmberg, D. V. D. Ville, and T. Lasser, *Biomed. Opt. Express* **8**, 5637 (2017).
6. U. J. Birk, A. Darrell, N. Konstantinides, A. Sarasa-Renedo, and J. Ripoll, *Appl. Opt.* **50**, 392 (2011).
7. A. C. Kak and M. Slaney, *Principles of Computerized Tomographic Imaging* (Society for Industrial and Applied Mathematics, 2001).
8. B. W. Lindsey and J. Kaslin, *Zebrafish* **14**, 574 (2017).
9. J. R. Walls, J. G. Sled, J. Sharpe, and R. M. Henkelman, *Phys. Med. Biol.* **50**, 4645 (2005).
10. J. Michalek, *Microsc. Microanal.* **21**, 1602 (2015).
11. V. C. Torres, C. Li, W. Zhou, J. G. Brankov, and K. M. Tichauer, *Appl. Opt.* **60**, 135 (2021).
12. G. C. Antonopoulos, D. Pscheniza, R.-A. Lorbeer, M. Heidrich, K. Schwanke, R. Zweigerdt, T. Ripken, and H. Meyer, in *Three-Dimensional and Multidimensional Microscopy: Image Acquisition and Processing XXI* 8949, 894907 (2014).
13. S. Doran, K. Koerkamp, M. Bero, P. Jenneson, E. Morton, and W. Gilboy, *Phys. Med. Biol.* **46**, 3191 (2001).
14. K. Islam, J. Dempsey, M. Ranade, M. Maryanski, and D. Low, *Med. Phys.* **30**, 2159 (2003).
15. M. Haidekker, *Comp. Meth. Prog. Biom.* **80**, 225 (2005).
16. A. Marcos-Vidal and J. Ripoll, *Opt. Lasers Eng.* **135**, 106191 (2020).
17. F. Natterer, *The Mathematics of Computerized Tomography* (Society for Industrial and Applied Mathematics, USA, 2001).
18. V. Koljonen, O. Koskela, T. Montonen, A. Rezaei, B. Belay, E. Figueiras, J. Hyttinen, and S. Pursiainen, *Phys. Med. Biol.* **64**, 045017 (2019).
19. J. R. Walls, J. G. Sled, J. Sharpe, and R. M. Henkelman, *Phys. Med. Biol.* **52**, 2775 (2007).
20. O. Koskela, T. Montonen, B. Belay, E. Figueiras, S. Pursiainen, and J. Hyttinen, *Sci. Rep.* **9**, 13934 (2019).
21. A. Hendriksen, D. Schut, W. J. Palenstijn, N. Viganò, J. Kim, D. Pelt, T. van Leeuwen, and K. J. Batenburg, *Opt. Express* **29**, 40494 (2021).
22. W. van Aarle, W. J. Palenstijn, J. Cant, E. Janssens, F. Bleichrodt, A. Dabravolski, J. D. Beenhouwer, K. J. Batenburg, and J. Sijbers, *Opt. Express* **24**, 25129 (2016).
23. M. Grass, T. Köhler, and R. Proksa, *Phys. Med. Biol.* **45**, 329 (2000).
24. L. A. Feldkamp, L. C. Davis, and J. W. Kress, *J. Opt. Soc. Am. A* **1**, 612 (1984).
25. Y. Liu, J. Dong, C. Schmidt, A. Boquet-Pujadas, J. Extermann, and M. Unser, “Dataset and scripts used in ‘Artifacts in Optical Projection Tomography Due to Refractive-Index Mismatch: Model and Correction’,” Zenodo, 23 February 2022, <https://doi.org/10.5281/zenodo.6141020>.